

**977.** *Reactivity Differences between Hæmoglobins. Part IV.*<sup>1</sup>  
*The Thermodynamics of Ionization of Some Vertebrate Hæmoglobins.*

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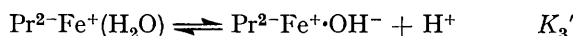
The ionization of eleven vertebrate methæmoglobins to their alkaline forms has been measured at various ionic strengths and temperatures. Large variations are observed in the enthalpy and entropy of ionization, but the free energy of ionization varies only slightly, owing to compensating changes in  $\Delta H$  and  $T\Delta S$ . It is concluded that the variations in  $\Delta H$  and  $T\Delta S$  are due to variations in electrostatic interactions originating from different charge configurations in each of the hæmoglobins. This conclusion is supported by the effect of ionic strength on the  $pK$  of ionization.

As early as 1928 Barcroft<sup>2</sup> drew attention to the fact that different vertebrate hæmoglobins have different affinities for oxygen and ascribed this to the differences in protein composition. The possible mechanisms by which changes in globin structure can alter the reactivity of a hæmoglobin at its iron atom have been discussed by us in Part I.<sup>3</sup> However, the problem remains of finding criteria by which the importance of each mechanism can be assessed. One such criterion is suggested from the analysis of the thermodynamics of ionization of the human methæmoglobins A, S, and C to their alkaline forms,<sup>3,4</sup> and it is the application of this criterion to the ionization of twelve other vertebrate hæmoglobins that concerns us in this Paper.

In Part I we concluded that the different entropies and enthalpies of ionization of human methæmoglobins A, S, and C may be ascribed to different electrostatic interactions in each hæmoglobin. In Part II<sup>4</sup> we employed Kirkwood's<sup>5</sup> and Tanford and Kirkwood's<sup>6</sup> theory of electrostatic interactions in globular proteins to show that the observed linearity of the plot of  $T\Delta S^\circ$  against  $\Delta H^\circ$  is consistent with this conclusion. In Part III<sup>1</sup> we extended the theoretical treatment given in Part II to include hæmoglobins which differ considerably in their charged amino-acids, and we showed that if for a series of methæmoglobins the plot of  $T\Delta S$  against  $\Delta H$  of ionization is linear, and the points for human methæmoglobins A, S, and C fall on the line, then: (i) the differences between the entropies and enthalpies of ionization are entirely electrostatic in origin; and (ii) the charge changes which differentiate the hæmoglobins do not occur at positions closer than 10 Å to the iron atoms.

In the present study we have measured the ionization of eleven vertebrate hæmoglobins at different temperatures and ionic strengths. We shall discuss these data in relation to the ideas developed in Parts II and III.

*The Variation of  $pK_3'$  with Temperature at Low Ionic Strength.*—Adopting the same nomenclature as in Part I, the ionization of a methæmoglobin to its alkaline form is represented by the equation



where  $\text{Pr}^2\text{-Fe}^+(\text{H}_2\text{O})$  and  $\text{Pr}^2\text{-Fe}^+\cdot\text{OH}^-$  are the acidic and alkaline forms respectively of methæmoglobin, and  $K_3'$  is the ionization constant at finite ionic strength.

The measurement of  $pK_3'$  for the methæmoglobins was made at different temperatures at ionic strength,  $I = 0.05$ . This value of  $I$  was chosen for two reasons: (1) The desirability of working at ionic strengths low enough to make valid the simplification to the

<sup>1</sup> Part III, Beetlestone and Irvine, preceding Paper.

<sup>2</sup> Barcroft, "The Respiratory Function of the Blood. Part II. Hæmoglobin," Cambridge University Press, London, 1928, ch. 5.

<sup>3</sup> Beetlestone and Irvine, *Proc. Roy. Soc.*, 1964, *A*, **277**, 401.

<sup>4</sup> Beetlestone and Irvine, *Proc. Roy. Soc.*, 1964, *A*, **277**, 414.

<sup>5</sup> Kirkwood, *J. Chem. Phys.*, 1934, **2**, 351.

<sup>6</sup> Tanford and Kirkwood, *J. Amer. Chem. Soc.*, 1957, **79**, 5333.

Kirkwood theory used in Part III. (2) The accuracy of the determination of  $pK_3'$  decreases below  $I = 0.05$  owing to small amounts of the ionizations characterized by  $pK_1'$  and  $pK_2'$  (see Part I).

The lowest ionic strength at which the effects of  $pK_1'$  and  $pK_2'$  on the spectrum are negligible was therefore chosen, namely  $I = 0.05$ . The data are summarized in Table 1. We determined  $\Delta H$  in the usual way from the plot of  $pK_3'$  against  $1/T$ . For all the haemoglobins we obtained good linear plots, some of which are shown as examples in Fig. 1.

TABLE 1.

Values of  $pK_3'$  at various temperatures and  $I = 0.0506$ .

Species	$pK_3'$ ( $I = 0.0506$ )			$\Delta G$	$\Delta H$	$T\Delta S$
Mouse (1) .....	8.273 (5.7)	8.148 (12.1)	8.000 (20.0)	10,726	7140	3590
Rat (2) .....	8.174 (4.2)	8.095 (12.2)	8.036 (20.0)	10,766	3340	7430
Baboon (3) .....	8.290 (6.0)	8.220 (12.0)	8.112 (20.0)	10,877	4660	6220
Patas monkey (4) .....	8.335 (5.2)	8.236 (12.2)	8.130 (20.0)	10,901	5180	5720
Mona monkey (5) .....	8.362 (5.2)	8.268 (11.7)	8.163 (20.0)	10,945	4930	6025
Pig (6) .....	8.484 (6.7)	8.396 (11.8)	8.211 (19.7)	11,009	7870	3140
Dog (7) .....	8.508 (5.3)	8.403 (12.4)	8.266 (20.0)	11,083	6090	4990
Hyaena (8) .....	8.490 (6.8)	8.415 (12.1)	8.295 (20.0)	11,122	5500	5620
Tantalus monkey (9) ...	8.535 (4.5)	8.420 (11.4)	8.300 (20.0)	11,129	5650	5480
Cat (10) .....	8.468 (5.5)	8.388 (11.4)	8.301 (20.0)	11,130	4350	6780
Pigeon (11) .....	8.602 (7.0)	8.494 (11.9)	8.290 (20.0)	11,115	9160	1960
Horse <sup>7</sup> (12) .....				11,276	3660	7620
Human A <sup>3</sup> (13) .....				10,914	3400	7510
Human S <sup>3</sup> (14) .....				10,986	4090	6900
Human C <sup>3</sup> (15) .....				10,928	4880	6050

The temperature at which each  $pK_3'$  was measured is indicated in parentheses beneath the value of the  $pK_3'$ . The values of  $\Delta G$ ,  $\Delta H$ , and  $T\Delta S$  are at  $20.0^\circ$ . The number in parentheses after each animal name gives the key to the points on the graphs in all the Figures. All values of  $pK_3'$  are subject to an error of between  $\pm 0.01$  and  $\pm 0.02$ .

$T\Delta S$  was calculated from the equation  $\Delta G = \Delta H - T\Delta S$ , using the values of  $\Delta G$  and  $\Delta H$  at  $I = 0.05$ . We did not attempt to calculate  $T\Delta S^\circ$ , the value of  $T\Delta S$  at  $I = 0$ , as is the usual practice. In the latter procedure  $\Delta G^\circ$  is obtained from  $pK^\circ$ , the value of  $pK$  extrapolated to zero ionic strength.  $\Delta H$  measured at some finite ionic strength is then assumed equal to  $\Delta H^\circ$ , the value of  $\Delta H$  at  $I = 0$ , and then  $T\Delta S^\circ = \Delta H^\circ - \Delta G^\circ$ . However, the assumption that  $\Delta H$  ( $I = 0.05$ ) is equal to  $\Delta H^\circ$  is not valid in these systems, since  $D_F$  and  $\partial(1/D_F)/\partial T$ , and consequently  $\Delta H$ , are functions of ionic strength. This is borne out by data at high ionic strength (unpublished results) which show  $\Delta H$  values significantly different from those at  $I = 0.05$ .

There are several outstanding features of the data in Table 1 and these will be considered separately. First, there is the variation in  $\Delta G$  as compared with that in  $\Delta H$  and in  $T\Delta S$ . The maximum variation in  $\Delta G$  is only 400 cal. mole<sup>-1</sup>, whereas the maximum variation in both  $\Delta H$  and  $T\Delta S$  is 6000 cal. mole<sup>-1</sup>. This is a consequence of the almost exact compensation of changes in  $\Delta H$  and  $T\Delta S$ , which is clearly demonstrated in Fig. 2, where  $T\Delta S$  is plotted against  $\Delta H$ . The slope of the line, as determined by a least-squares analysis, is  $0.995 \pm 0.025$ . A slope of unity corresponds to exact compensation of changes

<sup>7</sup> George and Hanania, *Biochem. J.*, 1953, **55**, 236.

in  $\Delta H$  and  $T\Delta S$ . The linearity of this plot, and the fact that its slope is close to unity, are both of interest, and we shall deal with these two points in turn.

Following the argument given in Part III, we conclude from the linearity of the plot of  $T\Delta S$  against  $\Delta H$ , and the fact that the points for methæmoglobins A, S, and C fall on the line, that (i) the differences between the  $T\Delta S$ 's and  $\Delta H$ 's of ionization are due to differences in electrostatic interactions in each of the hæmoglobins arising as a result of different charge distributions in each case; (ii) the charge changes which differentiate the hæmoglobins do not occur at positions closer than  $10 \text{ \AA}$  to the iron atoms. It seems reasonable from these observations to suggest that within the pH region 7.8–9.2, where the pK measurements are made, there is a charge configuration within  $10 \text{ \AA}$  of each of the iron atoms that is characteristic of a hæmoglobin, and that variations in charged amino-acids

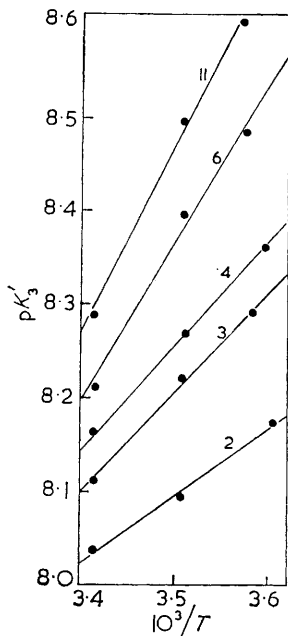


FIG. 1. Examples of the plots of  $pK'_3$  against  $1/T$  used to determine  $\Delta H$ . The numbers on the graph refer to the key given in Table 1.

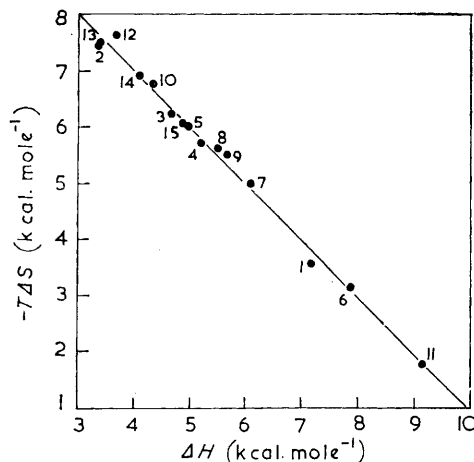


FIG. 2.  $T\Delta S$  against  $\Delta H$ , for the hæmoglobins in Table 1.

in this region would lead to drastic modification of the properties of hæmoglobin, for instance in the oxygen-binding capacity.

These conclusions depend only on the fact that the plot of  $T\Delta S$  against  $\Delta H$  is linear, and do not depend on the slope. This will depend on the ionic strength. Using the experimentally determined  $m_0$  for human methæmoglobins A, S, and C, calculation shows<sup>1</sup> that  $\bar{m}_0 = 1.16$  and  $\bar{m}_{0.05} = 1.08$ . This is to be compared with the value of  $\bar{m}_{0.05} = 0.995 \pm 0.25$  obtained in this work. The discrepancy between the calculated and experimental values of  $\bar{m}_{0.05}$  may be attributed to either: (a) the assumption that the  $\Delta H$ 's of ionization at  $I = 0.05$  for human methæmoglobins A, S, and C are equal to the  $\Delta H$ 's at  $I = 0$ , or (b) the assumption that simplifications made to the Kirkwood equations are strictly valid at  $I = 0.05$ .

A calculation of  $\bar{m}_{0.05}$  or  $\bar{m}_0$  from a knowledge of the dielectric constant of water and its temperature coefficient is not possible because, as Westheimer and Kirkwood<sup>8</sup> pointed out,

<sup>8</sup> Westheimer and Kirkwood, *Trans. Faraday Soc.*, 1947, **43**, 77.

the Kirkwood theory does not take into account electro-restriction of the solvent near charges. This limitation, in relation to the data on the ionization of human methæmoglobins A, S, and C, has been discussed by us in Part II. The question arises, however, as to whether the slope of the line in Fig. 2 is characteristic of hæmoglobins, or is characteristic of water as a solvent for reactions involving charged species. Data on the ionization of the bound water molecule in factor B of vitamin B<sub>12</sub> compared with those of the similar ionization in the mono-acid derivative, Factor V<sub>1A</sub>, favour the latter alternative (unpublished results). A preliminary survey of the available data on the association of small ions also confirms the view that  $m \approx 1$  is characteristic of water as a solvent for charged species. This argument will be developed in a later Paper.

We have mentioned earlier that the variation in  $\Delta H$  for the series of hæmoglobins is of the order of 6000 cal. mole<sup>-1</sup>. This large variation of  $\Delta H$  between different hæmoglobins has further significance. It is presumed in this study that the ionizing group is the same for all the hæmoglobins, the closely similar spectra of all the acid forms and all the alkaline forms making this a reasonable assumption. Thus we have the situation of a particular group (presumed to be the water bonded to the iron atom) ionizing with an enthalpy of ionization that varies between 3340 and 9160 cal. mole<sup>-1</sup>, depending on the charge environment. This observation shows the danger of attempting to identify ionizable groups in a protein solely by its enthalpy of ionization. Other workers<sup>9,10</sup> have also demonstrated the effect of environment on the thermodynamics of an ionizable group, and our present study confirms their observations.

*Effect of Ionic Strength.*—In Table 2 are recorded values of  $pK_3'$  for the different

TABLE 2.  
Values of  $pK_3'$  at various ionic strengths at 20.0°.

	Ionic strength, $I$					$-q$	$pK_3$	$\Delta G_{20}^\circ$ cal. mole <sup>-1</sup>
	0.0055	0.00928	0.0132	0.0174	0.0258			
Mouse .....	8.491	8.388			8.193	16.6 ± 0.8	9.22 ± 0.05	12,362
Rat .....	8.381	8.328	8.220		8.120	13.1 ± 0.5	8.98 ± 0.02	12,040
			(I = 0.0141)					
Baboon .....	8.417	8.357			8.222	11.1 ± 0.9	8.93 ± 0.04	11,973
Patas monkey ...	8.560		8.348	8.315	8.250	13.3 ± 0.9	9.06 ± 0.05	12,148
	(I = 0.0031)							
Mona monkey ...	8.350		8.295	8.265	8.230	6.5 ± 0.4	8.66 ± 0.03	11,611
Pig .....	8.650		8.478	8.450	8.316	15.1 ± 2.4	9.37 ± 0.07	12,563
Dog .....	8.572	8.500		8.407	8.356	10.3 ± 0.4	9.02 ± 0.02	12,094
	(I = 0.00716)							
Hyaena .....	8.682	8.583			8.435	13.8 ± 0.5	9.16 ± 0.03	12,282
Tantalus monkey	8.564	8.484			8.392	9.1 ± 0.4	8.98 ± 0.02	12,040
Cat .....	8.630	8.565			8.405	11.7 ± 0.6	9.03 ± 0.03	12,107
Pigeon .....	8.364	8.330			8.320	1.8 ± 0.6	8.48 ± 0.03	11,370
Horse <sup>7</sup> .....						7.8 ± 0.4	9.01 ± 0.02	12,082
Human <sup>8</sup> A .....						9.0 ± 1.0	8.81 ± 0.05	11,812
Human <sup>8</sup> S .....						7.5 ± 1.2	8.73 ± 0.06	11,709
Human <sup>8</sup> C .....						6.8 ± 0.5	8.66 ± 0.05	11,611

The values of  $pK_3'$  were obtained at the ionic strength indicated at the top of each column except where a different ionic strength is indicated in parentheses beneath the  $pK_3'$  value. Values of  $pK_3$  and  $q$ , the total charge on the protein, were obtained from plots of  $pK_3'$  against  $f(I)$ . All values of  $pK_3'$  are subject to an error of between ±0.01 and ±0.02.

hæmoglobins at various ionic strengths between 0.026 and 0.0055. We obtained  $pK_3$ , the value of  $pK_3'$  at zero ionic strength, by extrapolating to zero ionic strength the plot of  $pK_3'$  against  $f(I) = I^{1/2}/(1 + 10 I^{1/2})$ . The choice of this function for the hæmoglobin molecule has been discussed in Part I. The net charge,  $q$ , of a methæmoglobin in its acidic form is obtained from the graph by using the expression  $q = (S + A)/2A$  (see Part I), where  $S$  is the slope of the  $f(I)$  plot, and  $A$  is the Debye-Hückel coefficient. Values of  $q$ ,  $pK_3$ , and  $\Delta G^\circ$  for the different hæmoglobins are also recorded in Table 2.

<sup>9</sup> Hanania and Irvine, *J.*, 1962, 2745.

<sup>10</sup> Hanania and Irvine, *J.*, 1962, 2750.

Earlier in this Paper we concluded that the different thermodynamics of ionization of the various methæmoglobins originate from different charge effects. The different effects are now seen to be the result of differences in the number as well as in the configuration of the charges. For the human methæmoglobins A and S the difference between their free-energies of ionization (see Part II) is given by the equation

$$\Delta G_A^\circ - \Delta G_S^\circ = \frac{2N}{4.2 \times 10^7} \frac{e_q e_f}{r_{f,q} D_E} \text{ cal. mole}^{-1} \quad (2)$$

where  $e_q$ ,  $e_f$ ,  $r_{f,q}$ , and  $D_E$  have been defined in Part I. In general, for any hæmoglobin this equation is replaced by

$$\Delta G_A^\circ - \Delta G_x^\circ = \frac{N}{4.2 \times 10^7} \sum_{i=1}^{l=n} \frac{e_i e_f}{r_{i,f} D_{E(e,f)}} \text{ cal. mole}^{-1} \quad (3)$$

where  $r_{i,f}$  and  $D_{E(e,f)}$  are dependent on the relative positions of  $e_i$  and  $e_f$  and are defined in Part III. If a hæmoglobin differs from hæmoglobin A by  $n$  charges, and these occur at

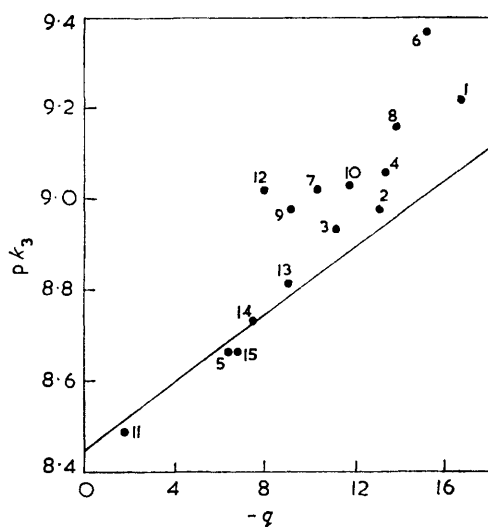


FIG. 3. Plot of  $pK_3$  against  $q$  for the fifteen hæmoglobins in Fig. 2. The line is the "theoretical" line of slope 0.0375 drawn through the points for human hæmoglobins A, S, and C.

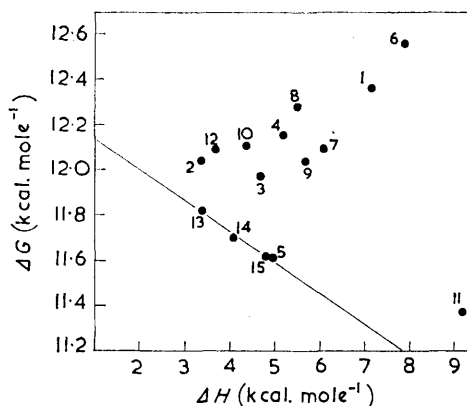


FIG. 4. Plot of  $\Delta G^\circ$  against  $\Delta H$  for the fifteen hæmoglobins in Fig. 2. The line is drawn through the points for human hæmoglobins A, S, and C.

positions such that for all these charges  $r_{i,f} = r_{f,q}$  and  $D_{E(e,f)} = D_E$ , then equation (3) becomes

$$\Delta G_A^\circ - \Delta G_x^\circ = \frac{nN}{4.2 \times 10^7} \frac{e_q e_f}{r_{f,q} D_E} \text{ cal. mole}^{-1} \quad (4)$$

Hence for hæmoglobins of this type a plot of  $\Delta G^\circ$  (or  $pK_3$ ) against  $q$  should be linear. Fig. 3 shows such a plot for the various vertebrate hæmoglobins. A "theoretical" line has been drawn through the points for hæmoglobins A, S, and C, the slope of which is based on the measured  $pK_3$  values and the known charge differences between hæmoglobins A, S, and C. The plotted points for hæmoglobins A, S, and C are those obtained when the value of  $q$  is derived from the plot of  $pK_3$  against  $f(I)$ , and, within experimental error, they fall on the "theoretical" line. The hæmoglobins of rat, mona monkey, and pigeon, within experimental error, also fall on the line, indicating that the charge changes which differentiate

them from hæmoglobin A are in similar positions to those which differentiate hæmoglobins S and C from A. That is to say, they are remote from all of the iron atoms.

All the other hæmoglobins are seen to fall above the line. This is a consequence of two factors. Firstly, the charge changes in hæmoglobins A, S, and C occur at positions remote from the four iron atoms, and hence for any hæmoglobin for which equation (3) rather than equation (4) is applicable, all the  $r_{i,f}$ 's must be less than  $r_{f,q}$ . This also implies a smaller value of  $D_E$ . Secondly, all the other hæmoglobins are more negatively charged than A, S, or C. Thus the value of  $\Delta G_A^\circ - \Delta G_S^\circ$  for these hæmoglobins must be greater than that for a hæmoglobin with the same number of charge changes but at distance  $r_{f,q}$ . Hence, in the plot of  $pK_3$  versus  $q$ , no points are found significantly below the line. There is an interesting corollary to the above, namely, that for a point to fall below the line the charge changes must occur nearer to the iron atom than those in A, S, and C and must be such as to make the net charge on the hæmoglobin more positive than on A.

The linearity of the plot of  $T\Delta S$  against  $\Delta H$  seems to imply that a plot of  $\Delta G$  against  $\Delta H$  should also be linear. However, this is not strictly true, as is evident from an inspection of Table I of Part II. This shows that whereas  $m_0$  (the slope of the plot of  $T\Delta S^\circ$  against  $\Delta H^\circ$ ) is constant to within  $\pm 5\%$  for  $45^\circ < \theta < 180^\circ$ ,  $m_0'$  (the slope of the graph of  $\Delta G^\circ$  against  $\Delta H^\circ$ , which is equal to  $1 - m_0$ ) varies by a factor of 2. Hence we cannot define an average  $m_0'$  that will remain constant for a series of hæmoglobins which differ by charges at many positions in the molecule. The large variation in  $m_0'$ , relative to that in  $m_0$ , is a consequence of the value of  $m_0$  being close to unity, and this in turn is due to the relative values of  $D_E$  and  $\partial(1/D_E)/\partial T$ , which are themselves functions of  $\theta$ . It is apparent that a plot of  $\Delta G^\circ$  against  $\Delta H^\circ$  will be linear only for hæmoglobins where  $D_E$  and  $\partial(1/D_E)/\partial T$  are strictly constant, and this situation is most closely approximated to when the charge changes which differentiate the hæmoglobins occur at the same point in the molecule. Thus the graph of  $\Delta G^\circ$  against  $\Delta H^\circ$  is linear for the human methæmoglobins A, S, and C.

To summarize, we conclude that the differences in the thermodynamics of ionization of the fifteen methæmoglobins studied are due to differences in the charge configuration on the globins, which arise from the various amino-acid replacements.

The question now arises whether these conclusions also apply to: (1) the reaction of methæmoglobins with other ligands over the entire pH range in which hæmoglobin is stable; (2) the ionization of hæmoglobins of animals classified as fish, amphibia, or reptiles; (3) the reactions of reduced hæmoglobin, in particular the oxygenation reaction. Experiments designed to answer these three questions are in progress.

## EXPERIMENTAL

*Methods.*—The procedure described in Part I was followed for determinations of  $pK_3'$ , except that a Pye Dynacap pH meter was used for the pH measurements. The manual temperature control was used.

*Material.*—The hæmoglobins were prepared by the method described in Part I. The animals have been referred to in the text by their common names. More precise names are as follows: Rat—*Wistar* strain *albino*; baboon—*Papio anubis*; patas monkey—*Erythrocebus patas*; tantalus monkey—*Cercopithecus aethiops*; mona monkey—*Cercopithecus mona*; pigeon—*Columbia livia*. Dog, pig, and cat were specimens of unknown types of domestic species. Mouse was from an unknown strain of laboratory mouse. Mouse and rat hæmoglobin was obtained from pooled samples, and the remainder from single animals. No tests were performed to establish the homogeneity of the samples. However the presence of more than one component in any hæmoglobin does not invalidate in any way the conclusions drawn in this Paper, since the measured  $pK_3$ 's are an average of those for the pure components.

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